Femtosecond Protein Nanocrystallography

CFEL

SCIENCE

Thomas White CFEL, DESY Hamburg

Collaborators

Henry N. Chapman^{1,2}, Petra Fromme³, Anton Barty¹, Thomas A. White¹, Richard A. Kirian⁴, Andrew Aquila¹, Mark S. Hunter³, Joachim Schulz¹, Daniel P. DePonte¹, Uwe Weierstall⁴, R. Bruce Doak⁴, Filipe R.N.C. Maia⁵, Andrew V. Martin¹, Ilme Schlichting^{6,7}, Lukas Lomb⁷, Nicola Coppola¹, Robert L. Shoeman⁷, Sascha W. Epp^{6,8}, Robert Hartmann⁹, Daniel Rolles^{6,7}, Artem Rudenko^{6,8}, Lutz Foucar^{6,7}, Nils Kimmel¹⁰, Georg Weidenspointner^{11,10}, Peter Holl⁹, Mengning Liang¹, Miriam Barthelmess¹², Carl Caleman¹, Sébastien Boutet¹³, Michael J. Bogan¹⁴, Jacek Krzywinski¹³, Christoph Bostedt¹³, Saša Bajt¹², Lars Gumprecht¹, Benedikt Rudek^{6,8}, Benjamin Erk^{6,8}, Carlo Schmidt^{6,8}, André Hömke^{6,8}, Christian Reich⁹, Daniel Pietschner¹⁰, Lothar Strüder^{6,10}, Günter Hauser¹⁰, Hubert Gorke¹⁵, Joachim Ullrich^{6,8}, Sven Herrmann¹⁰, Gerhard Schaller¹⁰, Florian Schopper¹⁰, Heike Soltau⁹, Kai-Uwe Kühnel⁸, Marc Messerschmidt¹³, John D. Bozek¹³, Stefan P. Hau-Riege¹⁶, Matthias Frank¹⁶, Christina Y. Hampton¹⁴, Raymond Sierra¹⁴, Dmitri Starodub¹⁴, Garth J. Williams¹³, Janos Hajdu⁵, Nicusor Timneanu⁵, M. Marvin Seibert⁵, Jakob Andreasson⁵, Andrea Rocker⁵, Olof Jönsson⁵, Stephan Stern¹, Francesco Stellato¹, Karol Nass², Robert Andritschke¹⁰, Claus-Dieter Schröter⁸, Faton Krasniqi^{6,7}, Mario Bott⁷, Kevin E. Schmidt⁴, Xiaoyu Wang⁴, Ingo Grotjohann³, James Holton¹⁷, Stefano Marchesini¹⁷, Raimund Fromme³, Sebastian Schorb¹⁸, Daniela Rupp¹⁸, Marcus Adolph¹⁸, Tais Gorkhover¹⁸, Martin Svenda⁵, Helmut Hirsemann¹², Guillaume Potdevin¹², Heinz Graafsma¹², Björn Nilsson¹² and John C. H. Spence⁴.

 Center for Free-Electron Laser Science, DESY, Notkestrasse 85, 22607 Hamburg, Germany.
University of Hamburg, Luruper Chaussee 149, 22761 Hamburg, Germany.
Department of Chemistry and Biochemistry, Arizona State University, Tempe, Arizona 85287-1604 USA.
Department of Physics, Arizona State University, Tempe, Arizona 85287 USA.
Laboratory of Molecular Biophysics, Department of Cell and Molecular Biology, Uppsala University, Husargatan 3 (Box 596), SE-751 24 Uppsala, Sweden.
Max-Planck Advanced Study Group, Center for Free Electron Laser Science (CFEL), Notkestrasse 85, 22607 Hamburg, Germany.
Max-Planck-Institut für medizinische Forschung, Jahnstr. 29, 69120 Heidelberg, Germany.
Max-Planck-Institut für Kernphysik, Saupfercheckweg 1, 69117 Heidelberg, Germany.
PNSensor GmbH, Otto-Hahn-Ring 6, 81739 München, Germany.
Max-Planck-Institut für extraterrestrische Physik, Giessenbachstrasse, 85741 Garching, Germany.
Photon Science, DESY, Notkestrasse 85, 22607 Hamburg, Germany.
LCLS, SLAC National Accelerator Laboratory, 2575 Sand Hill Road. Menlo Park, CA 94025, USA.
PULSE Institute and National Accelerator Laboratory, 2575 Sand Hill Road. Menlo Park, CA 94025, USA.
Lawrence Livermore National Laboratory, 7000 East Avenue, Mail Stop L-211, Livermore, CA 94551, USA.
Advanced Light Source, Lawrence Berkeley National Laboratory, Berkeley, California 94720, USA.
Institut für Optik und Atomare Physik, Technische Universität Berlin, Hardenbergstrasse 36, 10623 Berlin, Germany.



The PIs

Henry Chapman

Center for Free-Electron Laser Science, DESY Hamburg.

John Spence

Department of Physics, Arizona State University.

Petra Fromme

Department of Chemistry and Biochemistry, Arizona State University.

Ilme Schlichting

Max-Planck-Institut für medizinische Forschung, Heidelberg.

Janos Hadju

Laboratory of Molecular Biophysics, Department of Cell and Molecular Biology, Uppsala University.

Richard Neutze

Department of Chemistry, Biochemistry and Biophysics, University of Gothenburg.



The CAMP Team

The CAMP instrument was designed and commissioned by the Max Planck CFEL Advanced Study Group

Sascha Epp¹, Robert Hartmann^{1,2}, Daniel Rolles¹, Artem Rudenko¹, Lutz Foucar¹, Benedikt Rudek¹, Benjamin Erk¹, Carlo Schmidt¹, André Hömke¹, Nils Kimmel², Christian Reich², Günther Hauser², Daniel Pietschner², Peter Holl², Hubert Gorke³, Helmut Hirsemann⁴, Guillaume Potdevin⁴, Tim Erke⁴, Jan-Henrik Mayer⁴, Heinz Graafsma⁴, Michael Matysek⁵, Sebastian Schorb⁶, Daniela Rupp⁶, Marcus Adolph⁶, Tais Gorkhover⁶, Christoph Bostedt⁷, John Bozek⁷, Marc Messerschmidt⁷, Joachim Schulz⁴, Lars Gumprecht⁴, Andrew Aquila⁴, Nicola Coppola⁴, Frank Filsinger⁸, Kai-Uwe Kühnel⁹, Christian Kaiser⁹, Claus-Dieter Schröter⁹, Robert Moshammer⁹, Faton Krasniqi¹, Simone Techert^{1,10}, Georg Weidenspointer², Robert L. Shoeman¹¹, Ilme Schlichting^{1,11}, Lothar Strüder^{1,2} and Joachim Ullrich^{1,9}

Max Planck Advanced Study Group at CFEL, 22761 Hamburg, Germany.
Max Planck Halbleiterlabor, 81739 München, Germany.
FZ Jülich, 52428 Jülich, Germany.
Deutsches Elektronen Synchrotron, 22607 Hamburg, Germany.
Universität Hamburg, 22607 Hamburg, Germany.
Technische Universität Berlin, 10623 Berlin, Germany.
LCLS, Menlo Park, USA.
Fritz-Haber-Institut der MPG, Berlin, Germany.
Max-Planck-Institut für biophysikalische Chemie, 37077 Göttingen, Germany.
Max-Planck-Institut für medizinische Forschung, 69120 Heidelberg, Germany



Key Advantages

- Needs only small crystals (sub-micron sized).
- Can use radiation-sensitive samples.
 - Illumination is fast and crystals are not reused (serial crystallography). Specimen damage as it is conventionally understood is irrelevant.
- Extra information arises from the coherence of the beam across the entire crystal.
- Offers high time resolution when doing pumpprobe experiments, and can study irreversible reactions.



Big proteins...

Life relies on separating "inside" from "outside".



Membrane thickness: about 5-6 nm.

Image: http://en.wikipedia.org/wiki/File:Annular_Gap_Junction_Vesicle.jpg (public domain)



Big proteins...

Transmembrane proteins control (amongst many other things) what may cross the membrane.



Image: http://en.wikipedia.org/wiki/File:Polytopic_membrane_protein.png CC-BY-SA



Experimental Setup



LCLS, AMO beamline, 2 keV, 70 fs pulse duration.

11.

SCIENCE

Diffraction Patterns





The "Monte Carlo" Method

By measuring a dataset with very high redundancy (>1000 measurements per independent reflection), we get accurate intensities despite the lack of rotation:

Kirian et al., Optics Express 18 (2010) p5713-5723



Merging of Intensities

If the symmetry of the crystal comes through in the final results, things are probably not going too badly.



Merging of Intensities



Another way to evaluate the quality of the data is to compare against synchrotron data, but beware...



Phasing the Data

At this point, our data goes into the conventional MX analysis pipeline.





Some numbers from "3PCQ"...

- Number of crystals: 15,445
- **Crystal size**: 0.2 2 μm
- **Data frames per crystal:** 1
- **Temperature:** room temperature
- Hydration: Fully hydrated, in mother liquor
- **Exposure time:** 70 fs
- Number of images collected: ~ 1,800,000
- Oscillation angle: zero
- Data reduction software: CrystFEL
- X-ray energy (per exposure): random
- X-ray energy (mean): 2 keV
- X-ray bandwidth: ~ 0.1%

Anton Barty, CFEL



Software: CrystFEL

Suite of programs which share code and file formats, "CCP4 style".

- It's easy to interchange real and simulated input.
- Time consuming steps are multi-threaded.

SCIENCE



Reciprocal Space

Section through reciprocal lattice with Ewald sphere overlaid.





Reciprocal Space

Truncation of crystal lattice leads to "truncation rods" at each point of the reciprocal lattice.

Infinite lattice

Truncated lattice





Reciprocal Space





Peak Shapes - Rear Detector







Specimens

- Photosystem 1
- Lysozyme
- R. Viridis reaction centre (light/dark)
- Elastase
- **Proteinase K**
- Cathepsin B (glycosylated form)
- Photosystem 1-ferredoxin cocrystals (light/dark)
- Photosystem 2 (light/dark)
- Haemoglobin



The next year or two

- Develop analysis algorithms get even better results.
- New injector technology waste less sample.
- Use unique information (between Bragg peaks) to solve structures in new ways.
- Demonstrate conventional phasing methods: SAD/MAD etc.

... solve lots of otherwise inaccessible structures!



Looking ahead to 2015...

High repetition rate FELs, combined with detectors which can keep up, can make this go a **lot** faster...



Conclusions

- The feasibility of doing crystallography in the "diffract and destroy" regime using a femtosecond laser has been demonstrated.
- Even at this early stage, new structural information is being obtained.
- There are many more exciting things to try...



Main publication Chapman et al., Nature (2011) – out on the 3rd Feb Injector DePonte et al., J. Phys. D 41, 195505 (2008) **CAMP** instrument Strüder et al., Nuclear Instruments and Methods in Physics Research A 614 (2010) 483-496 Monte Carlo integration Kirian et al., Optics Express 18 (2010) 5713-5723 PDB entry 3PCQ Poster #130 (page 25), Koopmann et al. ... and many more in preparation ...!

SCIENC