

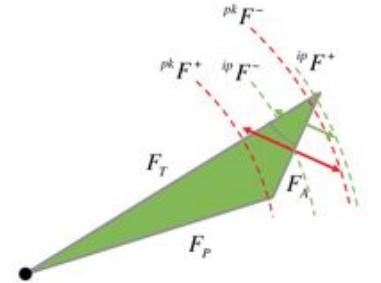
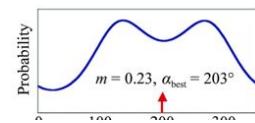
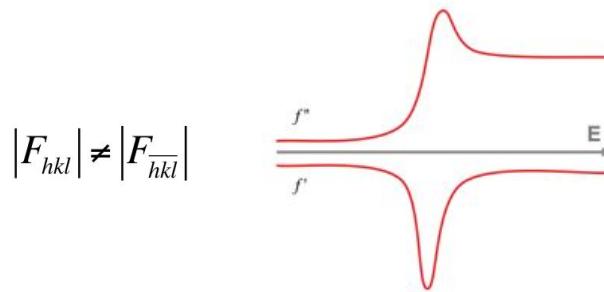
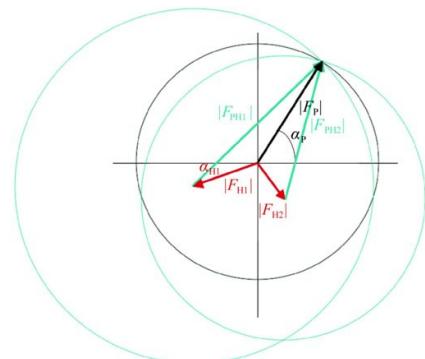
# Biology IV: Experiments / Refinement

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 10/7/2014  
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## Recap

- Phasing methods
  - Ab initio
  - Molecular Replacement – 25%. Model bias
  - Multiple Isomorphous Replacement
  - Multiple Anomalous Diffraction
  - Single Anomalous Diffraction – 1%. phase ambiguity
- Importance of accurate measures of data quality



# Today

- Experiments
  - Beams
  - Diffractometry
  - Low-energy data collection
  - Automation
- Refinement
- XFEL

## Diffraction angles and resolution

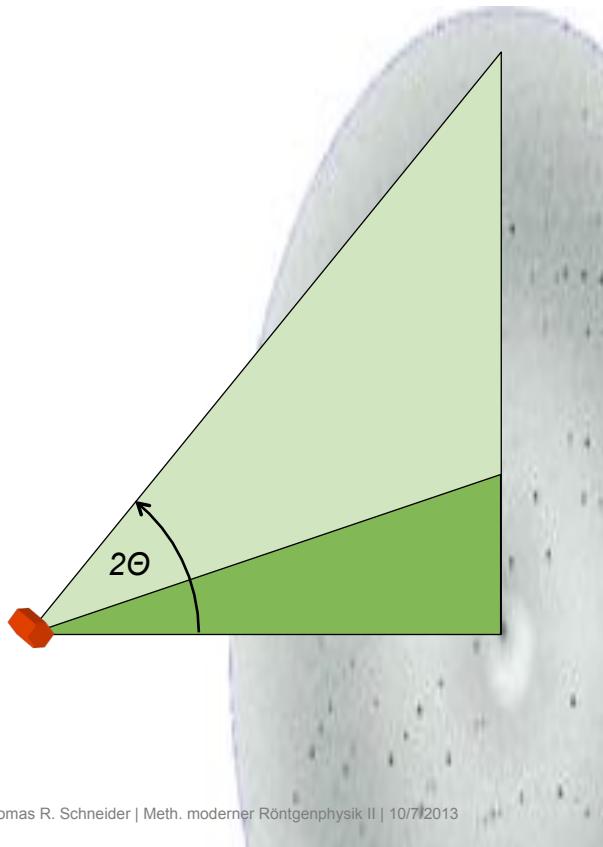
- Smaller lattice spacings  $d$  give rise to higher diffraction angles  $\Theta$ :

$$2d \sin\Theta = n\lambda$$

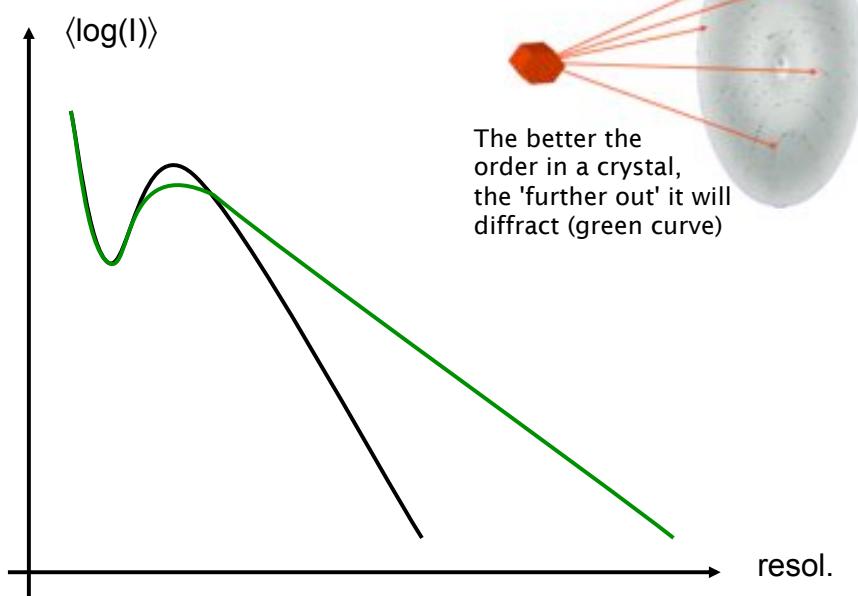
$$\Theta = \arcsin(n\lambda / 2d)$$

$$\begin{aligned}\arcsin(1\text{\AA}/2 * 3\text{\AA}) &= 9.6^\circ \\ \arcsin(1\text{\AA}/2 * 1\text{\AA}) &= 30.0^\circ\end{aligned}$$

- The lattice spacing roughly corresponds to the optical resolution.
- The higher the diffraction angle, the weaker is the average diffracted intensity

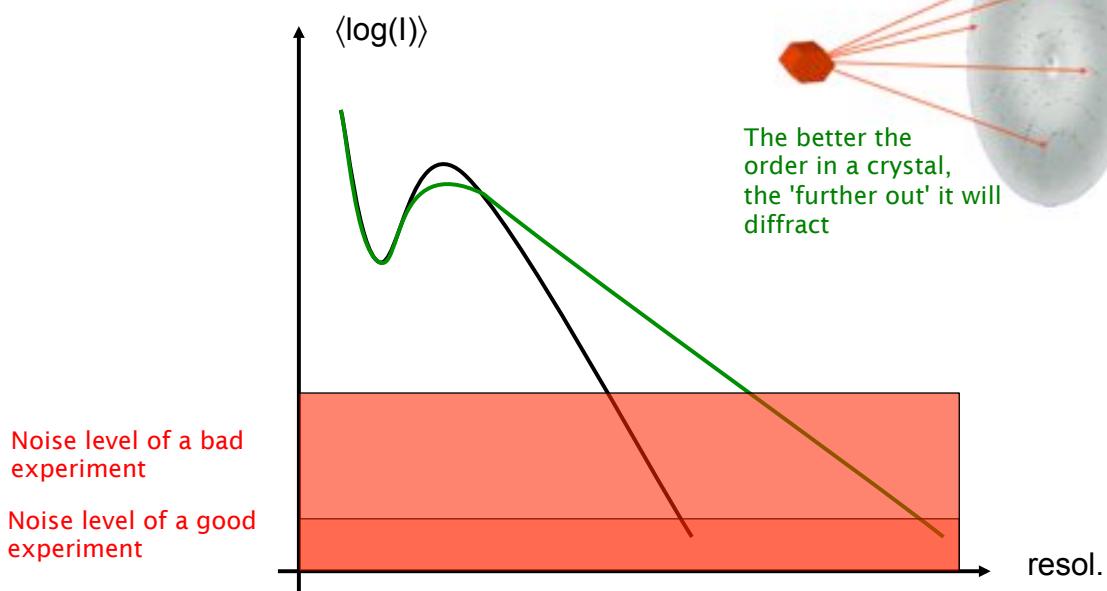


## Limited resolution in crystallography



- Diffraction data have an enormous dynamic range
- The higher the resolution / the larger the unit cell, the smaller the diffracted intensities

## Limited resolution in crystallography

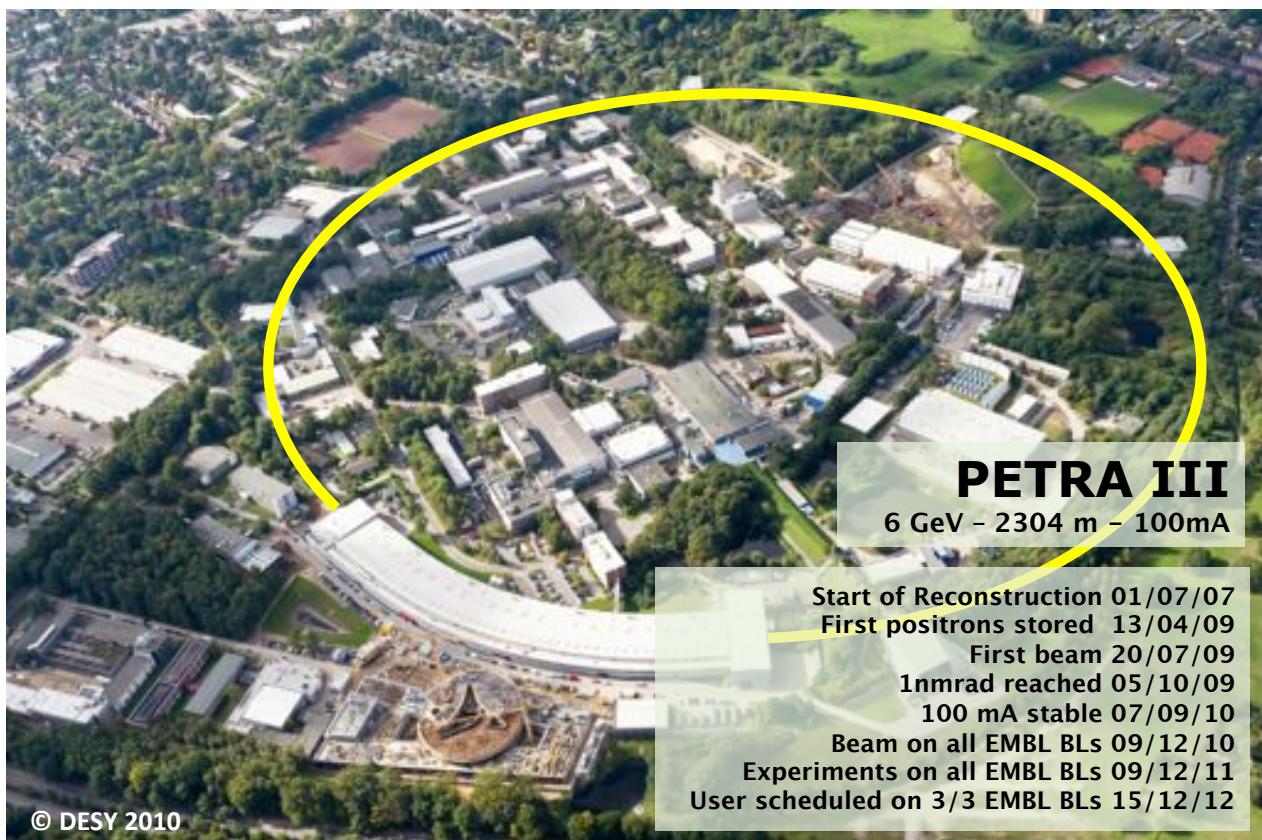


- Diffraction data have an enormous dynamic range
- The higher the resolution / the larger the unit cell, the smaller the diffracted intensities

## Why does anomalous phasing not work all the time?

- Molecule of interest does not contain anomalous scatterers
  - Seleno-Met incorporation
  - Derivatization
  - Co-crystallization
- The signal is too weak (small differences between large numbers)
  - Increase signal by choosing the right wavelength (synchrotron ...)
  - Enhance signal e.g. by oxidation of Se-atoms
  - Decrease background by optimizing the hardware, crystal mount, experiment
  - Add extra data -> SIRAS, MIRAS
  - More / more accurate data (inverse beam, kappa geometry)
- Crystals suffer from radiation damage
  - Keep crystal at cryogenic temperatures
  - Play tricks prolonging crystal lifetime (scavengers)
  - Careful use of tolerable dose (-> BEST)
  - Data from several positions on the same crystal
  - Data from several crystals

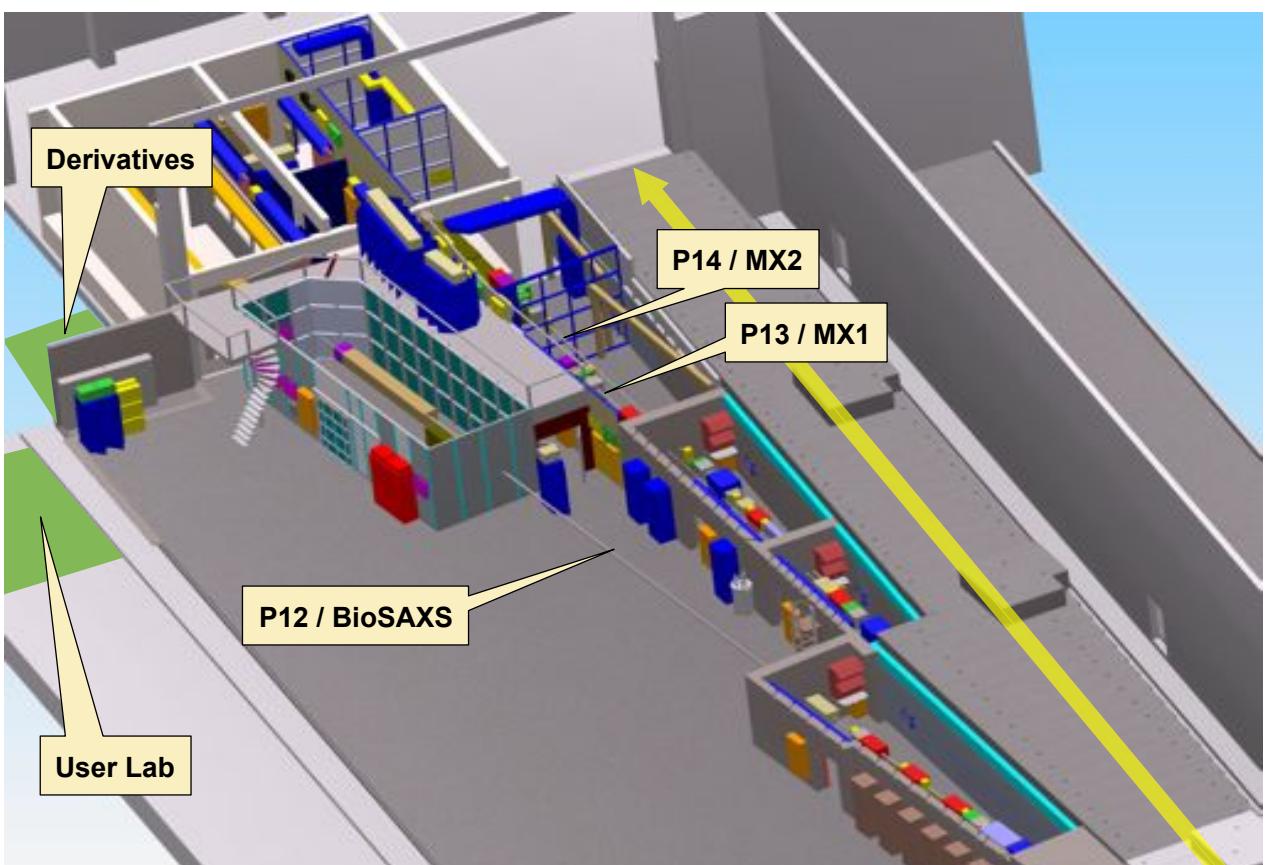
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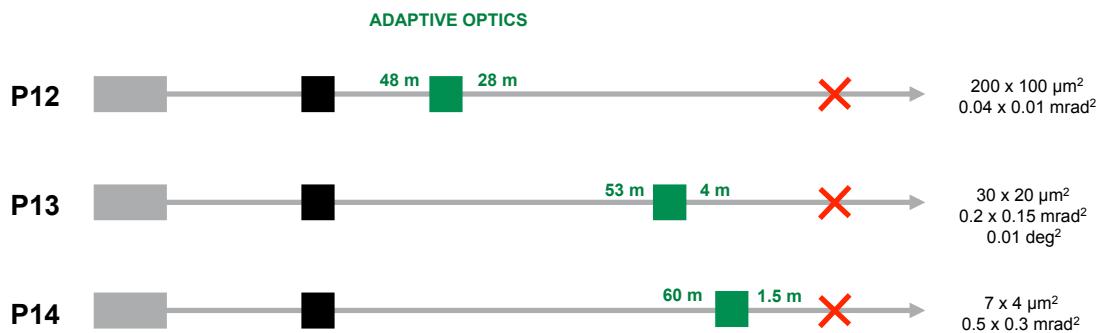


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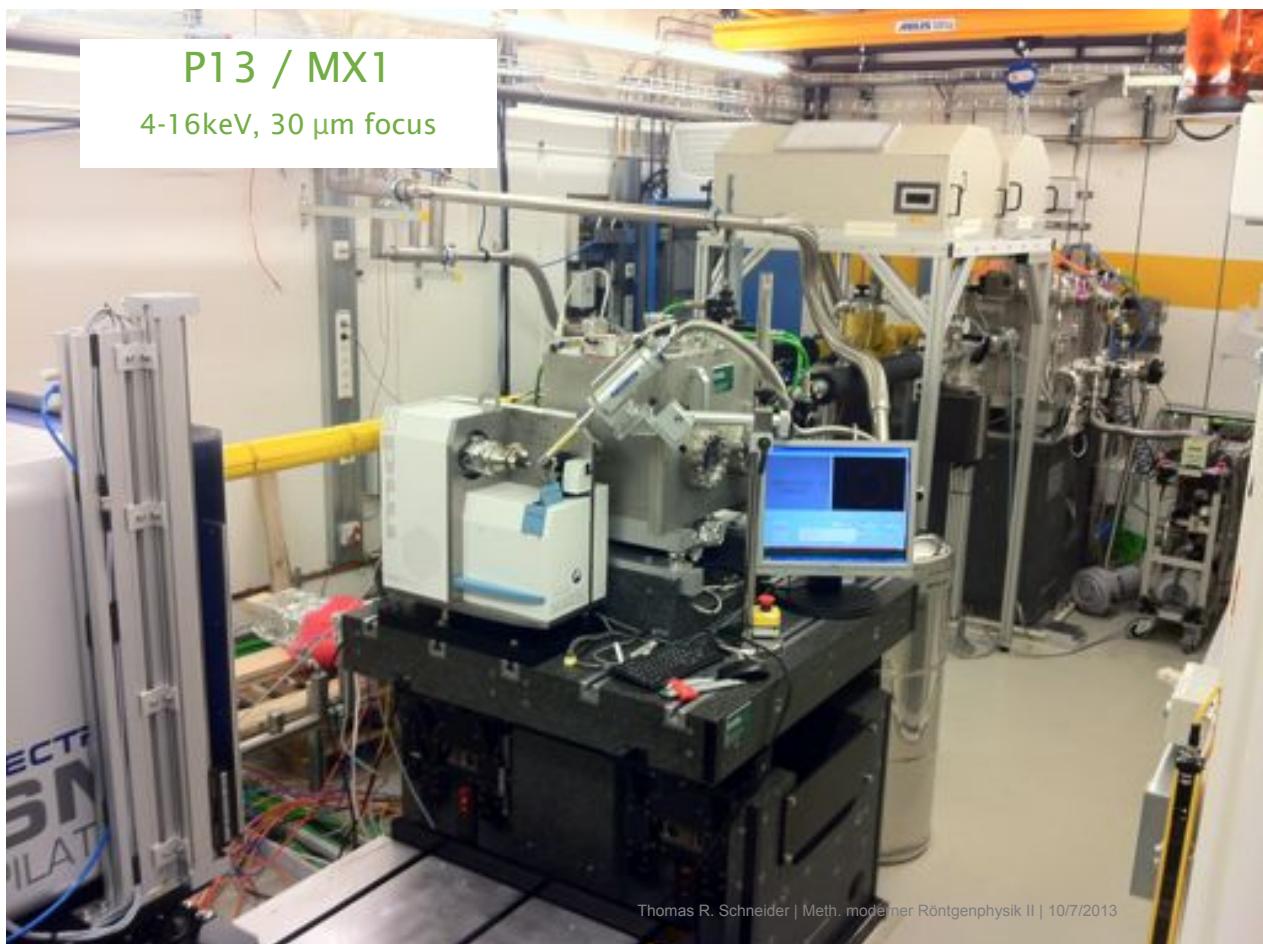
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## EMBL@PETRA3 - Optics



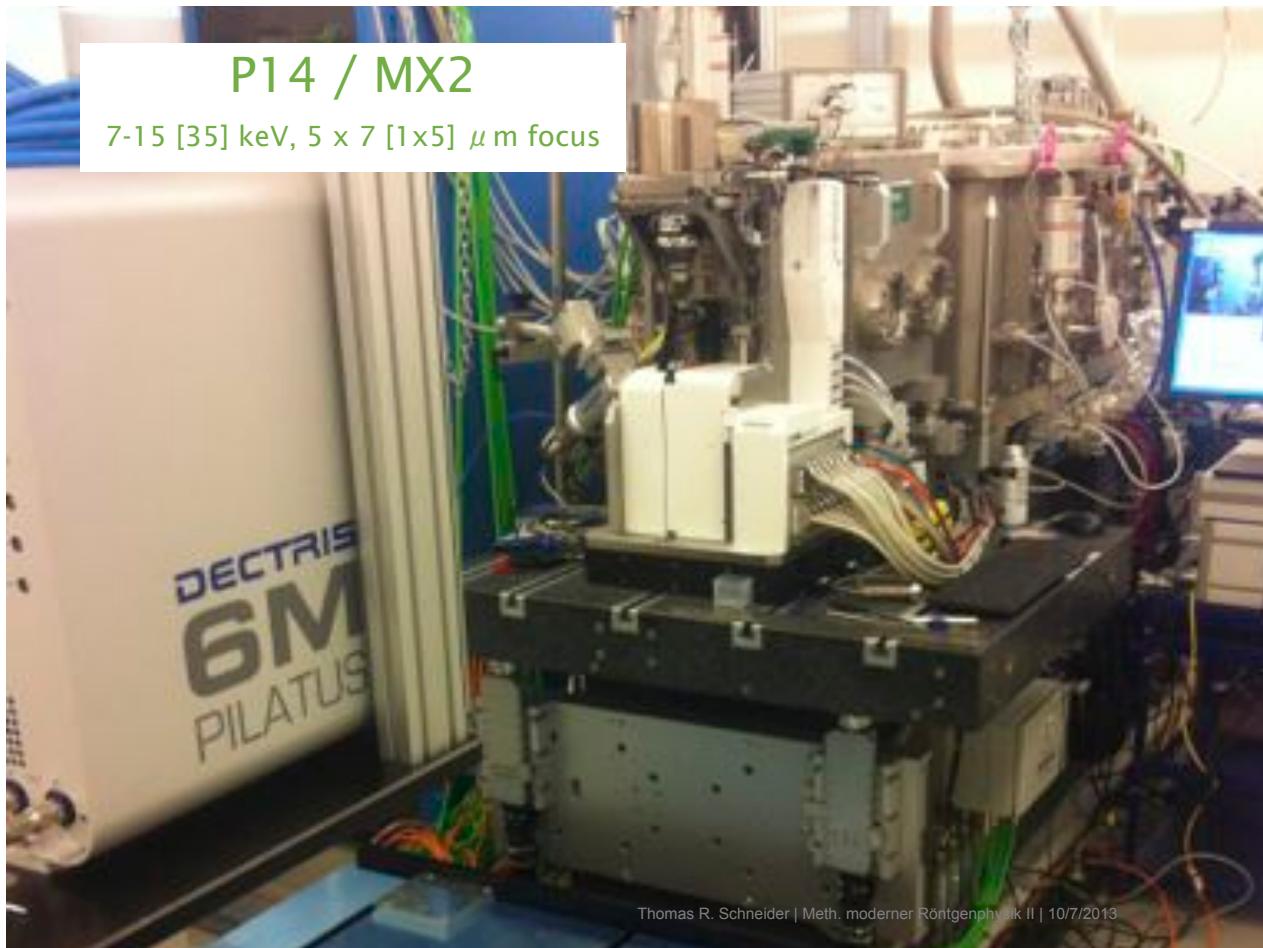
	P12:BioSAXS	P13:MX1	P14:MX2
Energy [keV]	4–20	5(4)–17	7–35
Monochromators	Si(111)	Si(111)	Si(111)
Beam size H x V [ $\mu\text{m}^2$ ]	$200 \times 60$	$28 \times 13$	$4 \times 1$
Divergence H x V [mrad]	$0.04 \times 0.01$	$0.2 \times 0.15$	$<0.5 \times <0.3$
Demagnification H / V	1:1.4 / 1: 1.2	1:12 / 1:15	1:60 / 1:40
Intensity [ph/sec]	$10^{13}$	$10^{13}$	$10^{12}$

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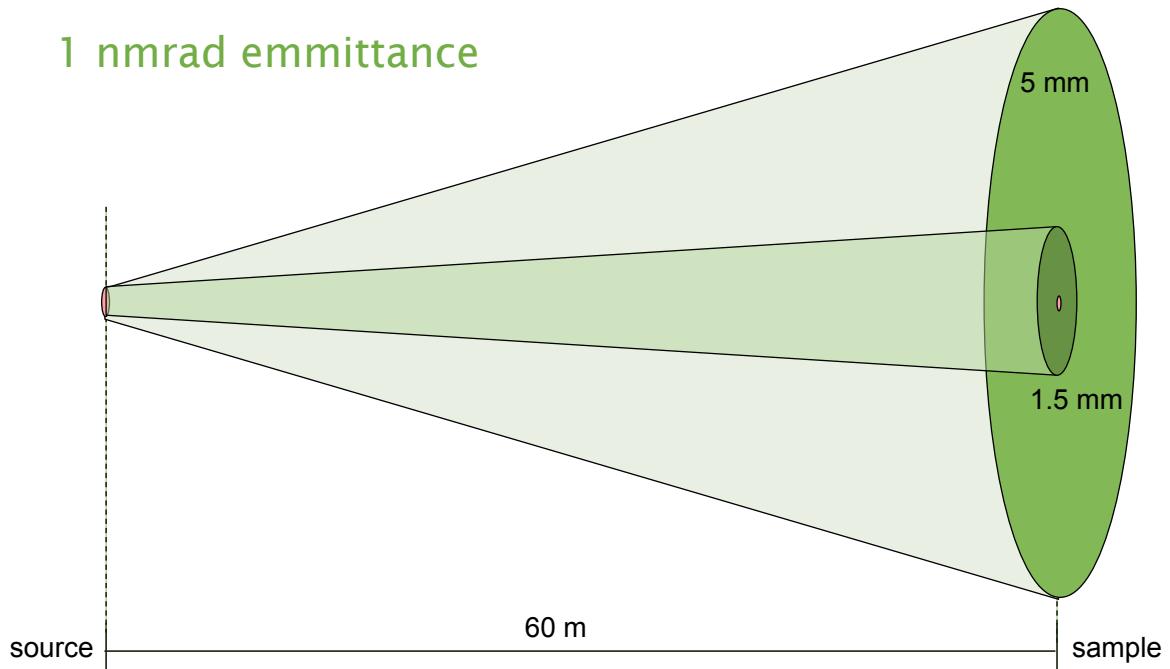


## P14 / MX2

7-15 [35] keV, 5 x 7 [1x5]  $\mu\text{m}$  focus



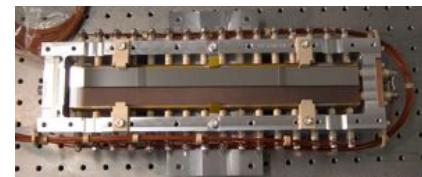
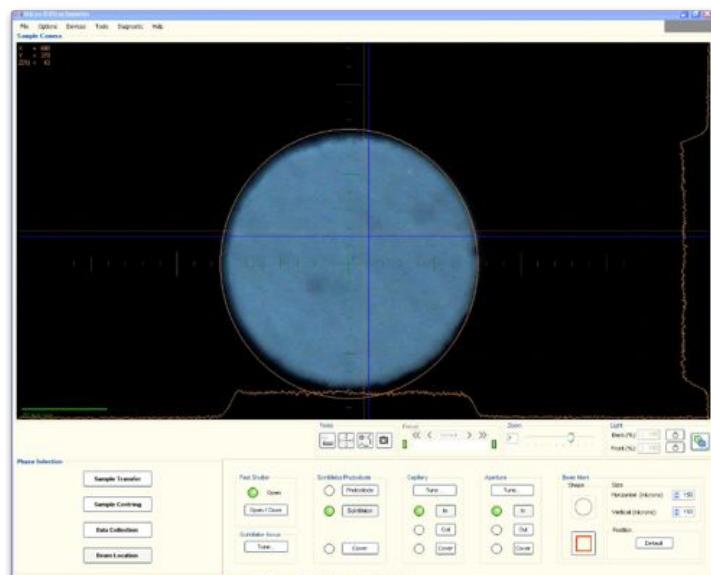
### 1 nmrad emittance



- $10^{13}$  monochromatic ph/sec to start with
- Flux through a 150  $\mu\text{m}$  aperture is ca.  $5 \times 10^{11}$  ph/sec  $\Rightarrow$  20 min. xtal-lifetime
- $20 \mu\text{rad} = 20 \mu\text{m}/\text{m}$ . Compare to  $0.1^\circ$  mos. =  $1700 \mu\text{rad}$

## P14: unfocused beam

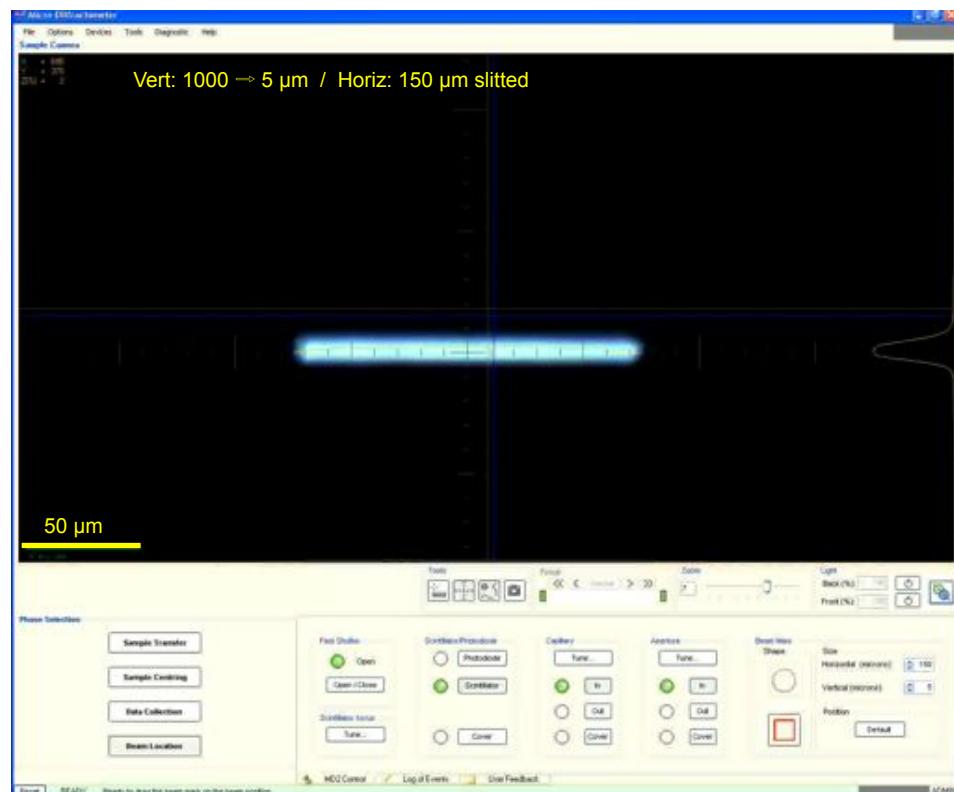
- Central beam profile is essentially top-hat
- flux:  $5 \times 10^{11}$  ph/sec in  $150 \mu\text{m} \varnothing$
- Beam position is very stable:
  - rmsd  $1.5 \mu\text{m}$  @  $< 10$  Hz
  - rmsd  $7 \mu\text{m}$  @  $> 30$  Hz
  - N.B. 60 m from the source, 16m from the DCM
- Introduce vertically focusing mirror to reduce beam size from  $1000 \mu\text{m}$  to  $5 \mu\text{m}$



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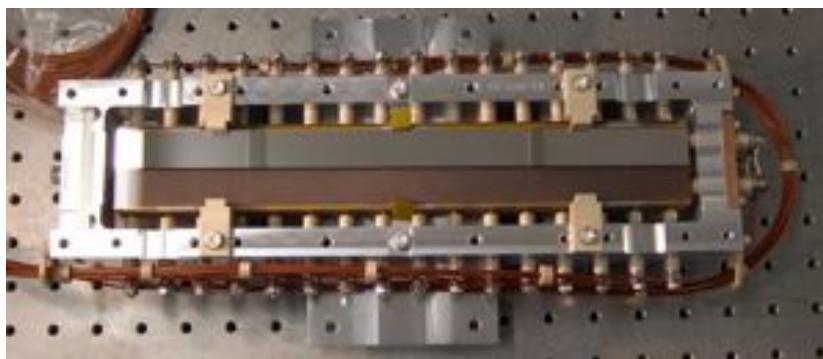
Vertical (Un)focusing ...



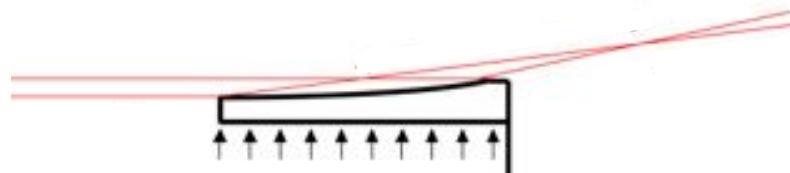
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## Adaptive X-ray focusing Mirrors

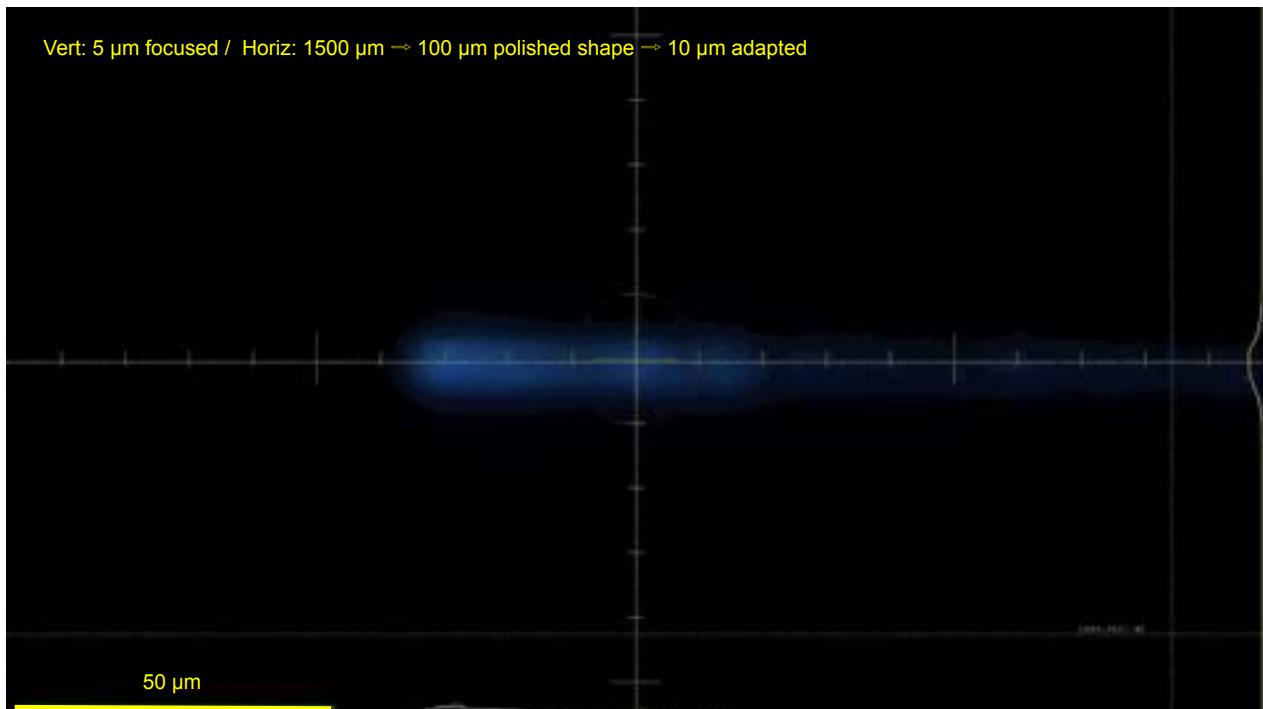


SESO bimorph mirror



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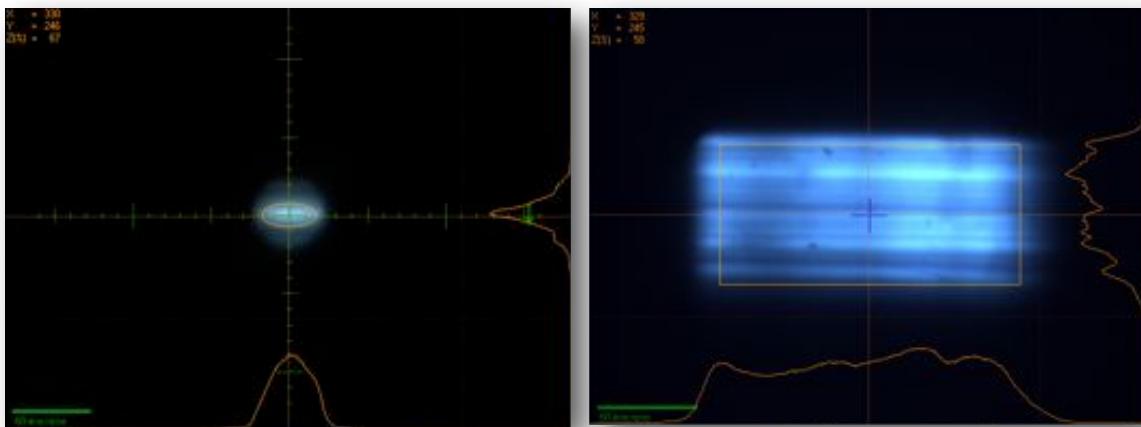
## P14: horizontal focusing (11/12/12)



- $10^{12}$  ph/sec into  $7 \times 4 \mu\text{m}^2$

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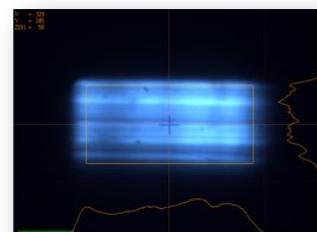
## P13: Focused beam I and II



- $H \times V$  (FWHM) =  $35 \times 15 \mu\text{m}^2$
- Focused, but detuned
- Shaped with Attocube-slits
- $H \times V$  (FWHM) =  $150 \times 70 \mu\text{m}^2$

## P13: Focused large beam, ConA [ 5/2/13]

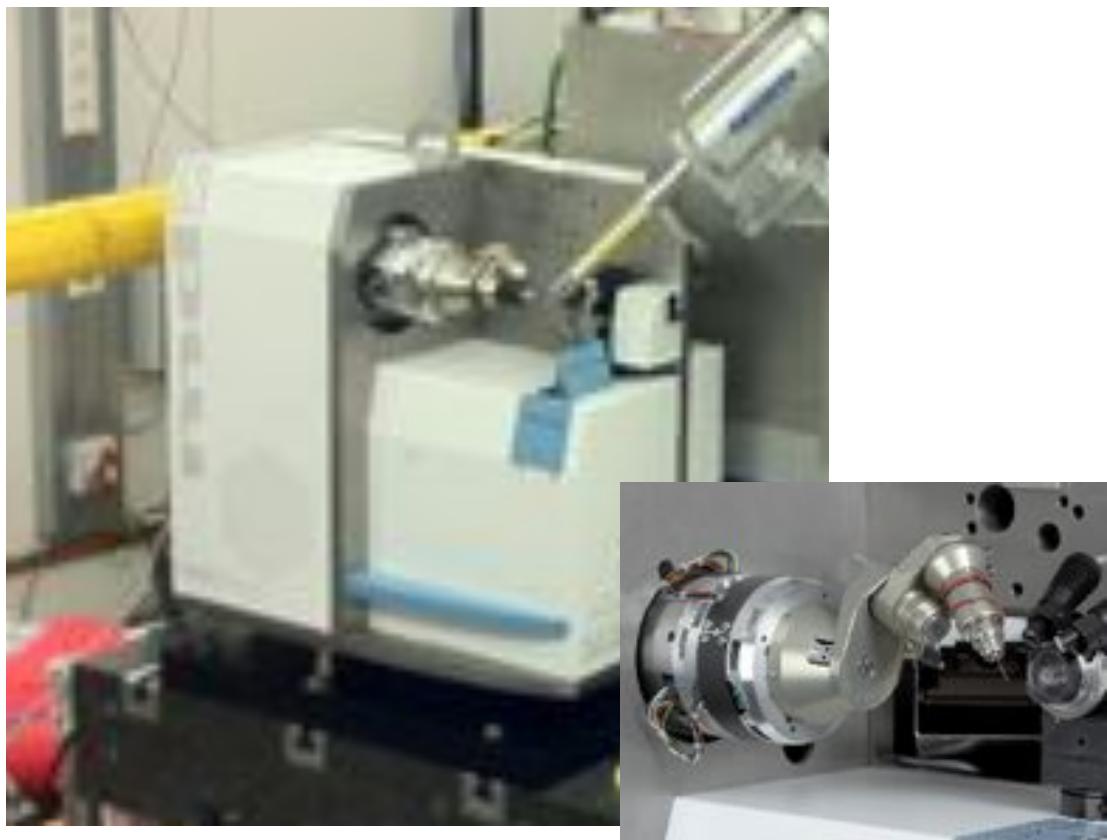
- Concanavalin A crystal  $\sim (50 \mu\text{m})^3$
- I222, 61.8 Å, 85.1 Å, 89.0 Å, 90°, 90°, 90°
- E = 15 keV,  $\lambda = 0.827 \text{ \AA}$ ,  $3 \times 10^{12} \text{ ph/sec}$  (no att.)
- 1600 frames @ 0.05 sec / 0.1° = 160° in 80 sec



SUBSET OF INTENSITY DATA WITH SIGNAL/NOISE >= -3.0 AS FUNCTION OF RESOLUTION													
RESOLUTION LIMIT	NUMBER OF OBSERVED	NUMBER OF UNIQUE	NUMBER OF POSSIB	COMPL OF DATA	R-FACTOR observed	R-FACTOR expected	I/SIGMA	R-meas	CC(1/2)	Anomal	SigAno	Nano	Corr
5.08	5743	1858	1876	99.0%	1.8%	1.8%	52.99	2.2%	99.9*	24*	0.974	718	
3.60	10368	3354	3373	99.4%	1.9%	2.0%	47.48	2.3%	99.9*	11*	0.900	1432	
2.94	13992	4371	4394	99.5%	2.8%	2.7%	35.16	3.3%	99.9*	7	0.853	1965	
2.54	15771	5159	5186	99.5%	4.5%	4.6%	21.05	5.5%	99.7*	-2	0.793	2262	
2.28	18306	5829	5874	99.2%	7.0%	7.0%	14.39	8.4%	99.4*	12*	0.849	2633	
2.08	20072	6463	6488	99.6%	9.2%	9.6%	10.54	11.1%	99.0*	1	0.782	2886	
1.92	21413	7019	7080	99.1%	13.7%	13.7%	7.27	16.6%	98.4*	2	0.808	3077	
1.80	23807	7521	7582	99.2%	25.5%	25.1%	4.26	30.7%	93.9*	-1	0.795	3416	
1.70	23621	7833	8113	96.5%	46.2%	44.9%	2.39	56.0%	78.7*	2	0.784	3386	
total	153093	49407	49966	98.9%	5.0%	5.0%	15.66	6.1%	99.9*	3	0.818	21775	

# Diffractometry

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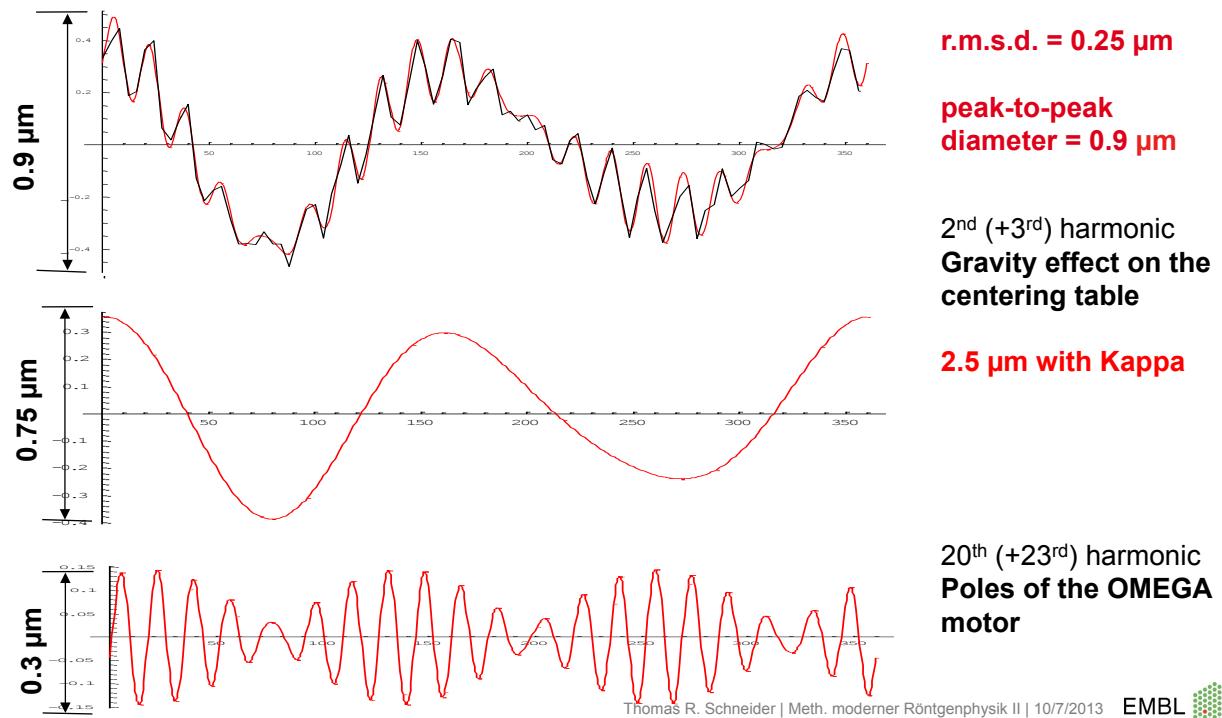


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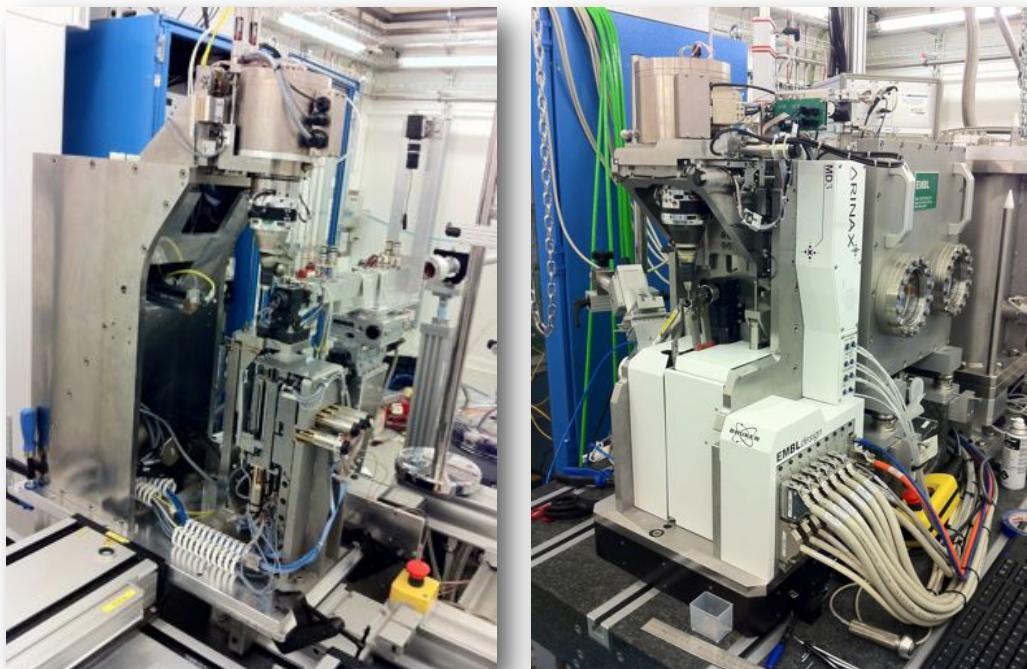
## MD2 - measured sample run out

Sample vs spindle position (video microscope + image processing), 1 Turn w/o Kappa

Black line – average of multiple measurements (varying conditions)  
Red lines – synchronous model



## Vertical Diffractometry

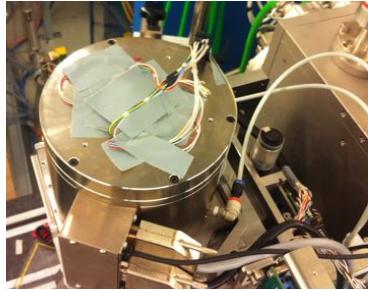


HPGonioV – summer 2011

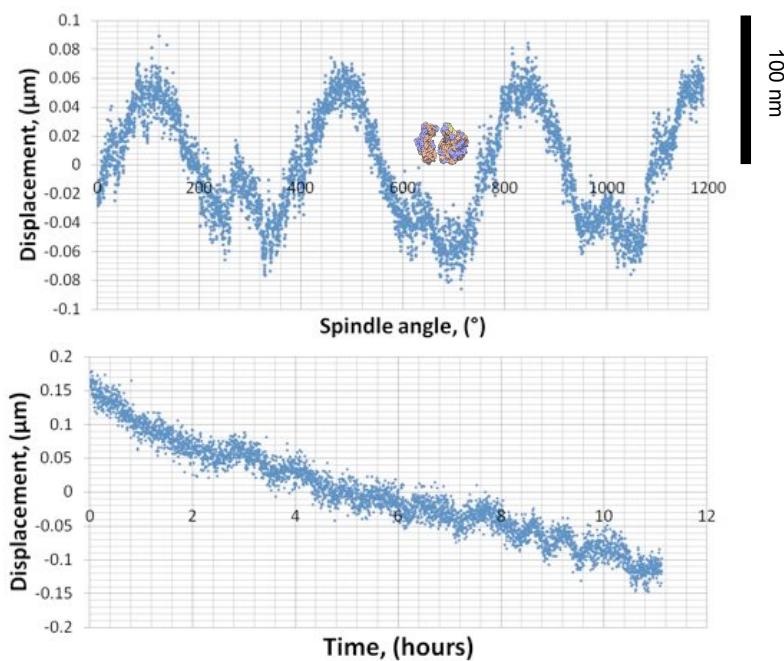
MD3 – 30/10/2012

## MD3 [ as of 19/01/2013]

- off-center component: 50 nm
- rmsd from ideal: 16 nm



- drift < 0.5 μm / 12 hrs  
(was > 10 μm before)



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## First data from MD3

Zn-free Insulin, 10 keV unfocussed, 5/11/12

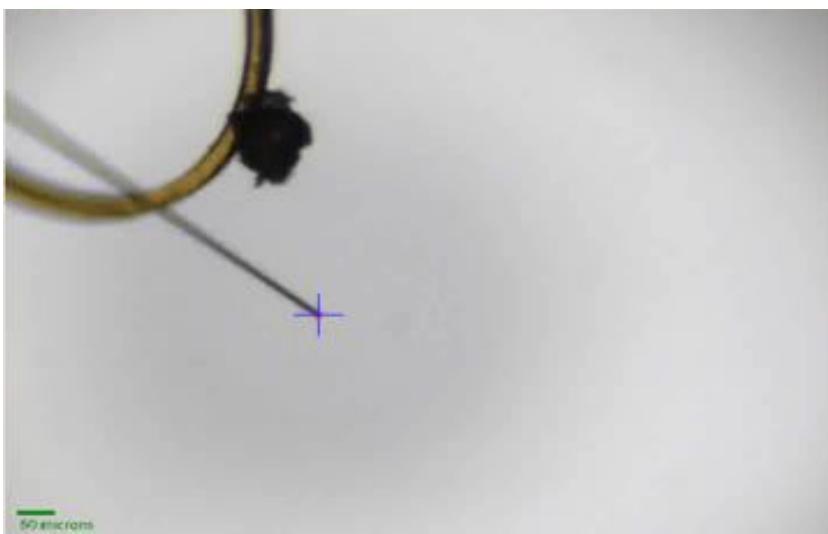
SUBSET OF INTENSITY DATA WITH SIGNAL/NOISE >= -3.0 AS FUNCTION OF RESOLUTION														
RESOLUTION LIMIT	NUMBER OF REFLECTIONS OBSERVED	NUMBER OF REFLECTIONS UNIQUE	NUMBER OF REFLECTIONS POSSIBLE	COMPLETENESS OF DATA	R-FACTOR observed	R-FACTOR compared expected	I/SIGMA	R-meas	CC(1/2)	Anomal Corr	SigAno	Nano		
4.80	14704	747	747	100.0%	1.9%	2.1%	14704	154.15	1.9%	100.0*	94*	3.780	321	
3.40	27439	1350	1350	100.0%	1.8%	2.1%	27439	153.49	1.9%	100.0*	82*	2.267	624	
2.78	36345	1753	1753	100.0%	2.1%	2.3%	36345	135.22	2.1%	100.0*	69*	2.103	827	
2.41	41700	2036	2036	100.0%	2.5%	2.7%	41700	110.24	2.6%	100.0*	66*	1.922	966	
2.16	47670	2363	2363	100.0%	3.2%	3.4%	47670	85.60	3.3%	100.0*	54*	1.578	1130	
1.97	51825	2565	2565	100.0%	4.4%	4.6%	51825	63.89	4.5%	100.0*	46*	1.394	1233	
1.82	56586	2789	2789	100.0%	7.0%	7.2%	56586	43.06	7.1%	99.9*	26*	1.122	1342	
1.70	60983	3051	3051	100.0%	11.0%	11.6%	60983	28.58	11.3%	99.8*	14*	0.970	1473	
1.61	44887	3061	3224	94.9%	17.6%	18.4%	44887	15.69	18.2%	99.2*	6	0.835	1453	
total	382139	19715	19878	99.2%	2.5%	2.7%	382091	71.28	2.6%	100.0*	36*	1.481	9369	

## Yb-loaded Thaumatin, 10 keV unfocussed, 6/11/12

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5.13	25296	1931	1931	100.0%	2.6%	2.7%	25296	89.08	2.7%	100.0*	98*	8.469	735	
3.64	47111	3491	3492	100.0%	2.5%	2.7%	47110	91.40	2.6%	100.0*	95*	5.418	1510	
2.97	61442	4521	4521	100.0%	2.8%	3.0%	61442	80.26	2.9%	100.0*	94*	4.770	2021	
2.58	71550	5335	5335	100.0%	3.7%	3.8%	71550	61.80	3.8%	99.9*	91*	4.172	2428	
2.30	81067	6034	6034	100.0%	4.7%	4.8%	81067	49.94	4.9%	99.9*	87*	3.453	2780	
2.10	90969	6729	6729	100.0%	5.8%	6.1%	90969	40.72	6.1%	99.9*	81*	2.696	3122	
1.95	98538	7278	7278	100.0%	7.9%	8.3%	98538	31.50	8.2%	99.9*	73*	2.138	3403	
1.82	105584	7822	7822	100.0%	12.0%	13.2%	105584	21.52	12.5%	99.7*	64*	1.712	3669	
1.72	74909	7658	8320	92.0%	16.5%	18.3%	74728	13.37	17.4%	99.0*	48*	1.285	3499	
total	656466	50799	51462	98.7%	4.1%	4.3%	656284	44.47	4.2%	100.0*	82*	3.032	23167	

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## '4D-Scan' (Florent Cipriani, Alexandre Gobbo, EMBL-GR)



- Synchronous movement of omega, centring table and alignment table using direct PMAC programming to collect data along a long needle in a 'continuous helical scan'
- Here: carbon fibre 10 micron thick, 650 micron long.
- Real movement is smooth. Jumps are due to recording

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## P14: 1D vs. 4D scan

- 'Monoclinic lysozyme' P2<sub>1</sub>  
31.5 53.0 72.0 90 99.2 90.0  
70% solvent for 129 residues.  
crystal 150 x 15 x ?? μm
- 10 keV = 1.24 Å
- half-focussed beam: 10 x 30 micron
- 1D: 0.2 deg / 0.04 sec, total = 36 secs.  
4D: 0.1 deg / 0.5 sec, total = 15 min.



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6.83	1422	427	432	98.8%	5.3%	5.3%	418	18.34	6.4%	99.4*	-11	0.760	238
4.85	2375	723	747	96.8%	7.5%	7.8%	2367	12.69	9.0%	99.1*	-6	0.769	352
3.97	3216	922	938	98.3%	6.6%	6.7%	3207	15.37	7.9%	99.3*	4	0.830	488
3.44	3764	1098	1122	97.9%	8.0%	8.1%	3752	12.68	9.5%	99.1*	-9	0.709	540
3.08	4341	1245	1264	98.5%	12.1%	12.0%	4328	8.93	14.3%	98.1*	-5	0.804	597
2.81	4774	1371	1379	99.4%	19.4%	19.3%	4767	5.93	23.0%	94.8*	-9	0.761	635
2.60	5122	1484	1503	98.7%	25.1%	25.3%	5106	4.44	29.8%	92.7*	-1	0.777	674
2.43	5442	1583	1611	98.3%	32.9%	32.0%	5430	3.59	39.1%	87.8*	-2	0.796	656
2.29	5068	1559	1707	91.3%	43.5%	41.1%	4942	2.61	52.2%	81.9*	-8	0.760	535
total	55464	10412	10703	97.3%	13.2%	13.1%	35317	7.75	15.7%	98.7*	-5	0.775	4715

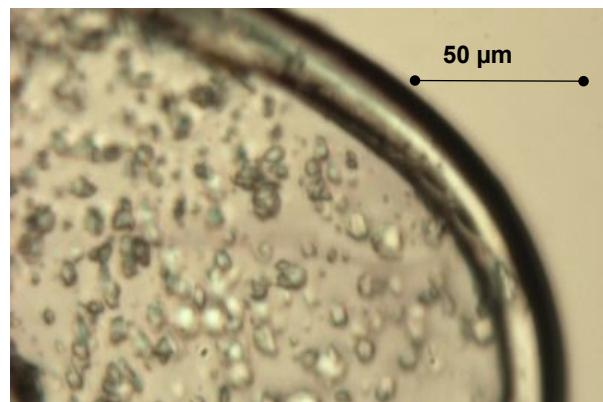
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4.77	3927	1208	1239	97.5%	4.0%	4.3%	3907	23.99	4.9%	99.5*	-17	0.701	576
3.38	7081	2107	2165	97.3%	4.3%	4.4%	7054	23.84	5.1%	99.6*	-16	0.744	964
2.76	9386	2737	2770	98.8%	5.7%	5.5%	9360	18.22	6.8%	99.4*	-15	0.763	1197
2.39	10840	3204	3272	97.9%	7.9%	7.8%	10799	13.02	9.4%	99.1*	-9	0.787	1285
2.14	12134	3623	3694	98.1%	11.6%	11.4%	12084	9.37	13.9%	98.1*	-4	0.784	1329
1.96	13387	3969	4078	97.3%	18.4%	19.1%	13328	6.03	21.9%	96.2*	-7	0.732	1404
1.81	1515	4346	4428	98.1%	35.3%	37.3%	14445	3.27	42.1%	88.9*	-9	0.682	1426
1.69	15419	4635	4725	98.1%	61.6%	66.7%	15338	1.89	73.5%	73.4*	-8	0.679	1430
1.60	10819	3945	5056	78.0%	82.1%	92.7%	10552	1.14	101.7%	52.5*	-9	0.645	791
total	97508	29774	31427	94.7%	8.3%	9.5%	96867	8.60	9.9%	99.6*	-9	0.728	10402

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## Microcrystal data collection

Ultralente Insulin in H3  
 $a=b=80 \text{ \AA}$   $c=37 \text{ \AA}$   
 crystal:  $(3\text{-}5) \times (3\text{-}5) \times (5\text{-}10) \mu\text{m}^3$   
 beam:  $4 \times 5 \mu\text{m}$  ('half'-focused)  
 exp: 1 s per  $1^\circ$  frame  
 ~40 frames/crystal  
 Pilatus6MF



### Partial data set statistics

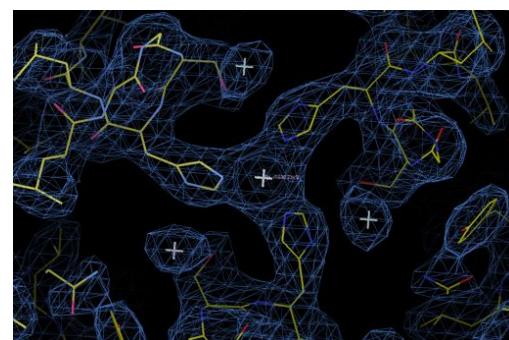
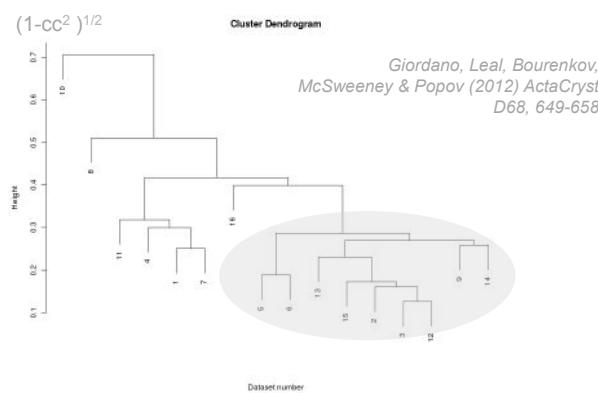
SUBSET OF INTENSITY DATA WITH SIGNAL/NOISE $\geq -3.0$ AS FUNCTION OF RESOLUTION											
RESOLUTION LIMIT	NUMBER OF REFLECTIONS OBSERVED	NUMBER OF REFLECTIONS UNIQUE	NUMBER OF REFLECTIONS POSSIBLE	COMPLETENESS OF DATA	R-FACTOR observed	R-FACTOR expected	R-FACTOR COMPARED I/SIGMA	R-meas	CC(1/2)	Anomalous Corr	
8.34	86	78	168	46.4%	3.3%	3.3%	16	15.89	4.7%	99.6*	0
5.28	272	253	478	52.9%	4.5%	3.8%	38	13.53	6.4%	98.7*	0
3.93	540	496	960	51.7%	3.2%	3.6%	88	13.06	4.5%	99.4*	0
3.27	662	599	1154	51.9%	5.2%	5.1%	126	9.47	7.3%	99.1*	0
2.95	587	543	1012	53.7%	9.5%	8.7%	88	6.10	13.4%	97.8*	0
2.65	591	531	1420	37.4%	12.8%	12.7%	120	4.46	18.1%	97.0*	0
total	2738	2500	5192	48.2%	4.5%	4.5%	476	8.99	6.3%	99.5*	56

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## Ultralente insulin microcrystal structure solution

### Isomorphous subset via cluster analysis



Electron density map, model phases  
REFMAC R/Rfree=0.18/0.24

### Combined data set statistics

SUBSET OF INTENSITY DATA WITH SIGNAL/NOISE $\geq -3.0$ AS FUNCTION OF RESOLUTION													
RESOLUTION LIMIT	NUMBER OF REFLECTIONS OBSERVED	NUMBER OF REFLECTIONS UNIQUE	NUMBER OF REFLECTIONS POSSIBLE	COMPLETENESS OF DATA	R-FACTOR observed	R-FACTOR expected	R-FACTOR COMPARED I/SIGMA	R-meas	CC(1/2)	Anomalous Corr	SigAno	Nano	
8.34	1989	168	168	100.0%	8.2%	9.3%	1987	25.91	8.6%	99.7*	61*	1.800	83
5.28	6054	486	486	100.0%	11.2%	11.5%	6053	21.13	11.7%	99.4*	52*	1.548	242
3.93	11427	964	964	100.0%	13.5%	12.5%	11424	19.50	14.1%	99.1*	30*	1.227	479
3.27	14393	1182	1182	100.0%	22.6%	21.8%	14393	12.74	23.6%	99.2*	16*	0.967	591
2.95	12132	987	988	99.9%	43.8%	48.5%	12132	7.79	45.7%	97.5*	14	0.840	493
2.65	12892	1341	1442	93.0%	68.5%	82.9%	12822	4.28	72.0%	89.8*	-2	0.708	617
total	58887	5128	5230	98.0%	17.4%	17.7%	58811	12.07	18.1%	99.5*	23*	1.012	2505

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## Refinement

A 'good' model should ...

- Agree with the data

$$R = \frac{\sum_{hkl} |F_{hkl}^{obs} - F_{hkl}^{calc}|}{\sum_{hkl} |F_{hkl}^{obs}|}$$

- Agree with what we know

## Progress of a Refinement

**#par** is number of parameter in model  
**#obs** is number of reflections used in refinement  
**Rw** is Rwork in percent  
**Rf** is Rfree in percent  
**Rd** is the difference between Rw and Rf in percent  
**CPU** is approximate CPU time [secs] normalized to a Pentium III running at 500 MHz

step	data used	#par	#obs	Rw	Rf	Rd	CPU
molecular replacement	15.0-4.0	3	775	44.5	-	-	30
rigid body	10.0-2.5	9	2997	46.4	47.4	1.0	140
first round	10.0-1.5	3887	13818	30.2	34.3	4.1	500
after first rebuild 10cgls	"	3735	"	26.2	31.4	5.2	360
SHELXWAT	"	4091	"	20.2	24.6	4.4	2000
bld + SHELXWAT	"	4231	"	18.7	22.8	4.1	2000
include all data 20cgls	10.0-1.1	4203	33993	19.1	21.7	2.6	1450
ANIS 20cgls	"	9453	"	16.0	19.2	3.2	2850
Rebuild 10 SHELXWAT	"	9557	"	13.4	15.8	1.8	1480
rebuild 10 cgls	"	10481	"	12.3	15.3	3.0	1610
rebuild 10 cgls	"	10819	"	12.1	15.1	3.0	1650
rebuild 10 cgls	"	10838	"	11.6	14.6	3.0	1800
rebuild 10 cgls	"	11494	"	11.3	14.4	3.1	1800
rebuild 10 cgls	"	11576	"	11.0	14.0	3.0	1800
rebuild 10 cgls	"	11774	"	10.7	13.8	3.1	1800
put hydrogens	10.0-1.1	11765	"	9.7	12.8	3.1	2500
include test set	10.0-1.1	11693	35786	9.8	-	-	2600

## Target Function

- Implements our idea of agreement between the data and the measurements. One possible form is:

$$E_{X-ray} = \sum_{hkl} \left| F_{hkl}^{obs} - k F_{hkl}^{calc} \right|^2$$

- One could replace  $F_{obs}$  by  $I_{obs}$ .
- The  $hkl$ -terms can be weighted by the uncertainties of the measurements:  $1/\sigma(F_{obs})$ .
- Other functions, such as the  $CC(F_{obs}, F_{calc})$ , can be used.

## Observables and Parameters

res	#r/a	parameters	d:p
3.0	2	x y z	0.6:1
2.5	4	x y z B?	1:1
2.0	8	x y z B	2:1
1.5	20	x y z B	5:1
1.1	50	x y z U <sub>11</sub> U <sub>22</sub> U <sub>33</sub> U <sub>12</sub> U <sub>13</sub> U <sub>23</sub>	5:1
0.9	90	x y z U <sub>11</sub> U <sub>22</sub> U <sub>33</sub> U <sub>12</sub> U <sub>13</sub> U <sub>23</sub>	10:1

Numbers are for a protein crystal with 40% solvent.  
Lysozyme (129 amino acids 1001 atoms) will produce  
ca. 32000 reflections to 2 Å resolution.

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## Stereochemical Restraints

Acta Cryst. (1991). A47, 392–400

### Accurate Bond and Angle Parameters for X-ray Protein Structure Refinement

BY RICHARD A. ENGH AND ROBERT HUBER

Max Planck Institut für Biochemie, D8033 Martinsried bei München, Germany

Table 1. CSD parametrization atom types

Atom type	Description
C	Carbonyl C atom of the peptide backbone
C5W*	Tryptophan C <sup>12</sup>
CW*	Tryptophan C <sup>12</sup> , C <sup>13</sup>
CP*	Phenylalanine C <sup>1</sup>
CY*	Tyrosine C <sup>1</sup>
CY2*	Tyrosine C <sup>4</sup>
C5*	Histidine C <sup>5</sup>
CN*	Neutral carboxylic acid group C atom
CH1E	Tetrahedral C atom with one H atom
CH2E	Tetrahedral C atom with two H atoms (except CH2P, CH2G)
CH2P*	Proline C <sup>1</sup> , C <sup>2</sup>
CH2G*	Glycine C <sup>2</sup>
CH3E	Tetrahedral C atom with three H atoms
CR1E	Aromatic ring C atom with one H atom (except CR1W, CRH, CRHH, CRH)
CR1W*	Tryptophan C <sup>12</sup> , C <sup>13</sup>
CRH*	Neutral histidine C <sup>11</sup>
CRHH*	Charged histidine C <sup>12</sup>
N	Peptide N atom of proline
NR	Unprotonated N atom in histidine
NP	Pyrrole N atom
NH1	Singly protonated N atom (His, Trp, peptide)
NH2	Double protonated N atom
NH3	Triply protonated N atom
NC2	Arginine N <sup>6</sup> , N <sup>7</sup>
O	Carbonyl O atom
OC	Carboxyl O atom
OH1	Hydroxyl O atom
S	S atom
SM*	Methionine S atom
SH1E	Singly protonated S atom

\* Atom types marked with an asterisk are new (non-XPLOR) types.

Table 2. Bond parameters

Bond type	$\sigma$	Bond length (Å)
C5W-CW	0.018	1.433
CW-CW	0.017	1.409
C-CH1E	0.021	1.525
CS-CH2E	0.014	1.497
C5W-CH2E	0.031	1.498
CP-CH2E	0.023	1.502
CY-CH2E	0.022	1.512
C-CH2E		
CN-CH2E		
C-CH2G		
C5W-CR1E		
CW-CR1E		
CW-CR1W		
CP-CR1E		
CY-CR1E		
CY2-CR1E		
CS-CR1H		
C5-CR1E		
CW-CW-NHI		
C-N		
C-NC2		
C5-NH1		
CW-NH1		

Table 3. Angle parameters

Angle type	$\sigma$	Angle (°)	Angle t
C5W-CW-CW	1.2	107.2	CH3E-C
CW-C5W-CH2E	1.4	126.8	CH3E-C
CP-CR1E	1.0	133.9	CH3B-C
CY-CR1E	1.0	118.8	C-CH2E
CY2-CR1E	1.0	122.4	C5-CH2
CS-CR1H	1.6	106.3	CF-CH2
C5-CR1E	1.5	107.4	C5W-CI
C-N	1.5	116.9	CY-CH2
CH1E-C-N	2.0	116.2	C-CH2E
CH1E-C-NH1	1.7	120.8	C-CH2C
CH1E-C-O	2.5	117.0	C-CH2C
CH2E-C5-CR1E	1.3	129.1	CHIE-C
CH2E-C5-CR1H	1.3	131.2	CHIE-C
CH2E-CF-CR1E	1.7	120.7	CHIE-C
CH2E-C5W-CR1E	1.5	126.9	CHIE-C
CH2E-CY-CR1E	1.5	120.8	CHIE-C
CH2E-C-N	1.4	118.7	CHIE-C

Bond lengths and angles should be consistent with these values ('Engh&Huber Dictionary') derived from small molecule structures. ITC-F contains an updated version (§18.3).

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## Enforcing stereochemistry

$$E_{bond} = \sum_{bonds} \frac{|d^{obs} - d^{EH}|^2}{\sigma_{d^{EH}}}$$

- $E_{bond}$  is zero for a perfect agreement between expected bond lengths and bond lengths measured in the model. The value of  $E_{bond}$  increases for increasing deviations.
- The weight for each bond-type depends on the spread observed in small molecule structures

## Enforcing stereochemistry

- Target functions can be defined for various geometrical properties:

$$E_a = \sum_a \frac{|a^{obs} - a^{ex}|^2}{\sigma_{a^{ex}}}$$

- And combined into an overall function:

$$E_{chem} = E_{bond} + E_{angle} + E_{chir} + \dots$$

## Combined Target Function

$$E_{tot} = E_{chem} + w_{X-ray} E_{X-ray}$$

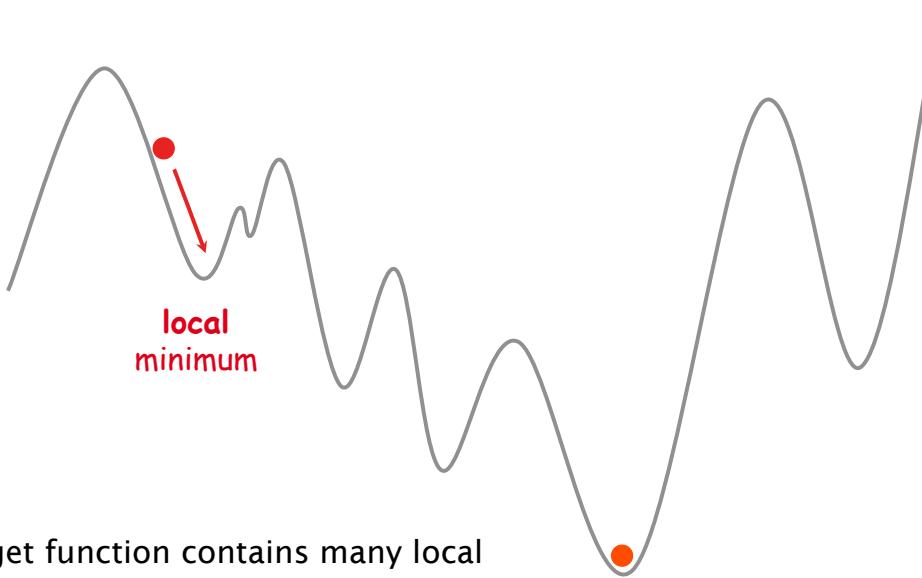
- $E_{chem}$  contains information about bond-lengths, - angles, planarity, chirality, non-bonded repulsion, electrostatic interactions etc. (the *a priori* knowledge).
- $w_{X-ray}$  is the relative weight between chemical information and experimental data. It depends not only on the amount of data but also on the quality of the data.

## Optimization of the Target Function

$$E_{tot} = E_{chem} + w_{X-ray} E_{X-ray}$$

- Minimization of  $E_{tot}$  should deliver a model that (1) is consistent with the data and (2) has reasonable stereochemistry.
- At 2 Å, 5000 atoms correspond to ca. 20000 parameters to be refined against 50000 reflections.

## The multiple minima problem

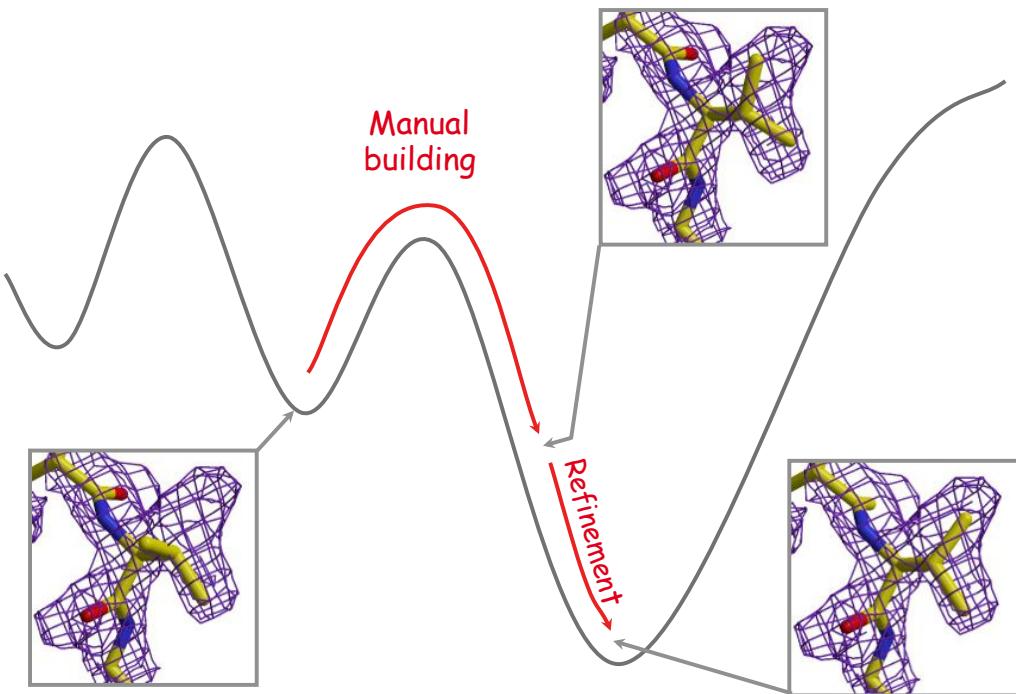


- The target function contains many local minima in addition to the global minimum
- Standard minimizers will reach the ‘closest’ local minimum but not the global minimum

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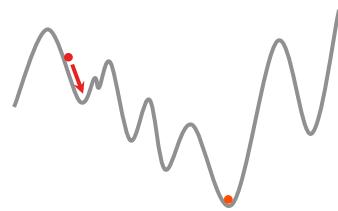
## Getting out of Local Minima



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## Simulated Annealing

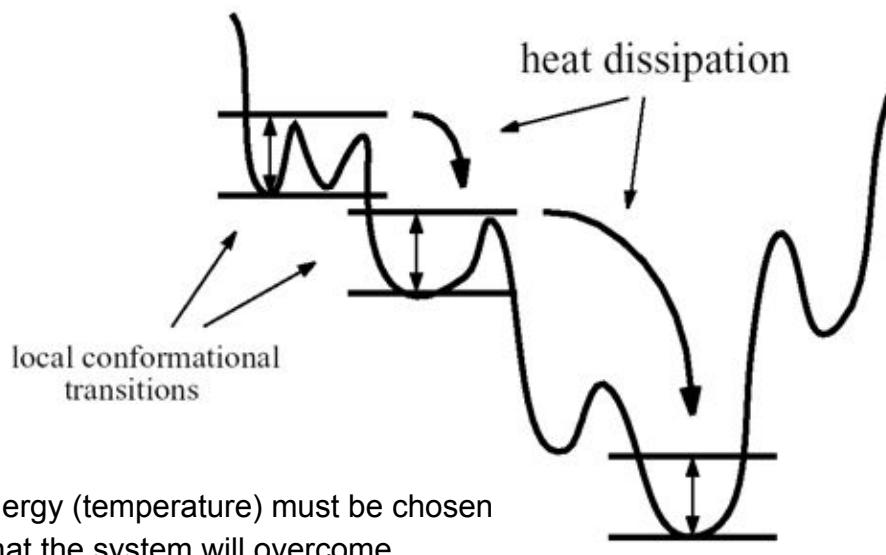


- A method to search conformational space
- *Annealing is a physical process wherein a solid is heated so that the particles assume random positions and then is cooled slowly so that all particles can arrange into their lowest energy state.*
- The target function  $E$  can be considered as the potential energy of the system and then a molecular dynamics simulation is run (in fact, historically, XPLOR (later CNS, later PHENIX) was an MD-program with an additional energy term  $E_{X-ray}$ ).

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## Simulated Annealing

Figure from  
Paul Adam's lecture notes



The energy (temperature) must be chosen such that the system will overcome barriers. But, once it arrives in the global minimum, it should not escape from there.

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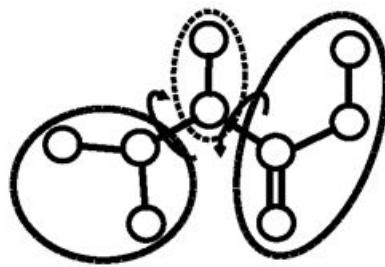
## Observables and Parameters

res	#r/a	parameters	d:p
3.0	2	x y z	0.6:1
2.5	4	x y z B?	1:1
2.0	8	x y z B	2:1
1.5	20	x y z B	5:1
1.1	50	x y z U <sub>11</sub> U <sub>22</sub> U <sub>33</sub> U <sub>12</sub> U <sub>13</sub> U <sub>23</sub>	5:1
0.9	90	x y z U <sub>11</sub> U <sub>22</sub> U <sub>33</sub> U <sub>12</sub> U <sub>13</sub> U <sub>23</sub>	10:1

## Constraints

- Constraints couple parameters to each other such that the overall number of parameters is reduced.
- Segmented rigid bodies
  - Entire residues
  - Rigid groups (guanodino, carboxylate, aromatic rings)
  - Entire domains

## Torsion Angle Refinement



- Deviations in bond lengths and angles are usually small.
- Torsion angles can be used as 'normal coordinates of protein dynamics'
- This reduces the number of parameters dramatically (up to a factor of 10)
- Initially introduced by Diamond (1971); implemented in XPLOR by Rice & Brunger (1996).
- Large radius of convergence (e.g. for bad molecular replacement models).

Diamond (1971) Acta Cryst. A27:436-452  
Sussmann (1977) Acta Cryst. A33:800  
Rice & Brunger (1996) Proteins 19:277-290

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## B-factors

- A 'crystal structure' represents a space and time average (many molecules in the crystal for the time of the experiment).
- The coordinates of an atom in the pdb-file represent the mean coordinates. The spread around the mean coordinates is described by the B-factor.
- Some Examples:

$B = 5 \text{ \AA}^2$	$\langle u \rangle = 0.25 \text{ \AA}$
$B = 20 \text{ \AA}^2$	$\langle u \rangle = 0.50 \text{ \AA}$
$B = 40 \text{ \AA}^2$	$\langle u \rangle = 0.71 \text{ \AA}$
$B = 80 \text{ \AA}^2$	$\langle u \rangle = 1.01 \text{ \AA}$

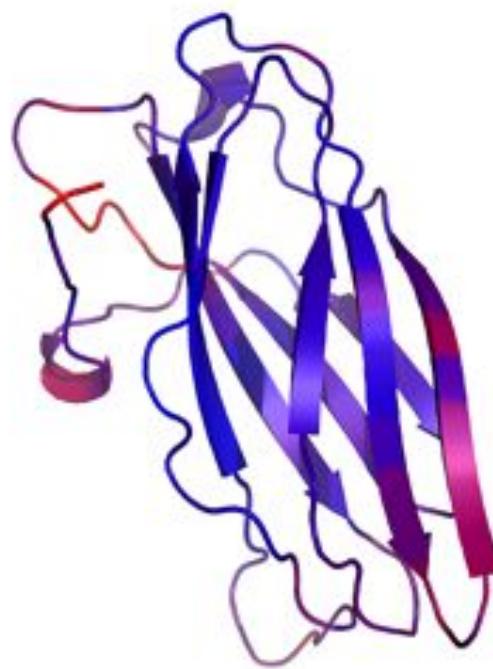
- A typical bondlength is 1.5 Å !

$$B = 8\pi^2\langle u^2 \rangle,$$

where  $\langle u^2 \rangle$  is the mean square displacement

## B-factors

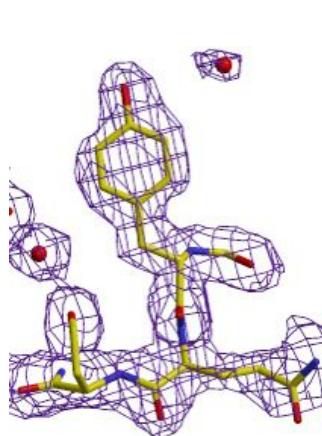
- Low
  - In well ordered region
- High
  - In loops
  - On the surface
  - In terminal regions
- Here
  - Red is high
  - Blue is low



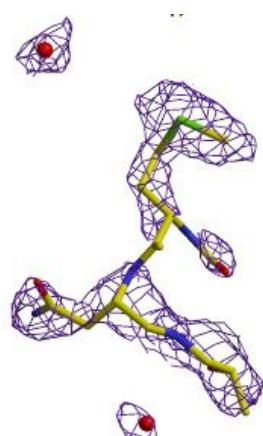
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## B-factors and electron density

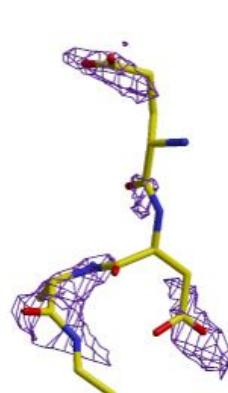
All pictures were taken from the same structure refined at 2.0 Å resolution.  
Countoured at the same level



Tyr48  
 $B = 20.9 \text{ \AA}^2$   
 $\text{RSR} = 0.12$



Met285/Asn286  
 $B = 69.9 \text{ \AA}^2 / 69.1 \text{ \AA}^2$   
 $\text{RSR} = 0.25 / 0.18$



Glu229/Asp230  
 $B = 73.9 \text{ \AA}^2 / 73.4 \text{ \AA}^2$   
 $\text{RSR} = 0.39 / 0.27$

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